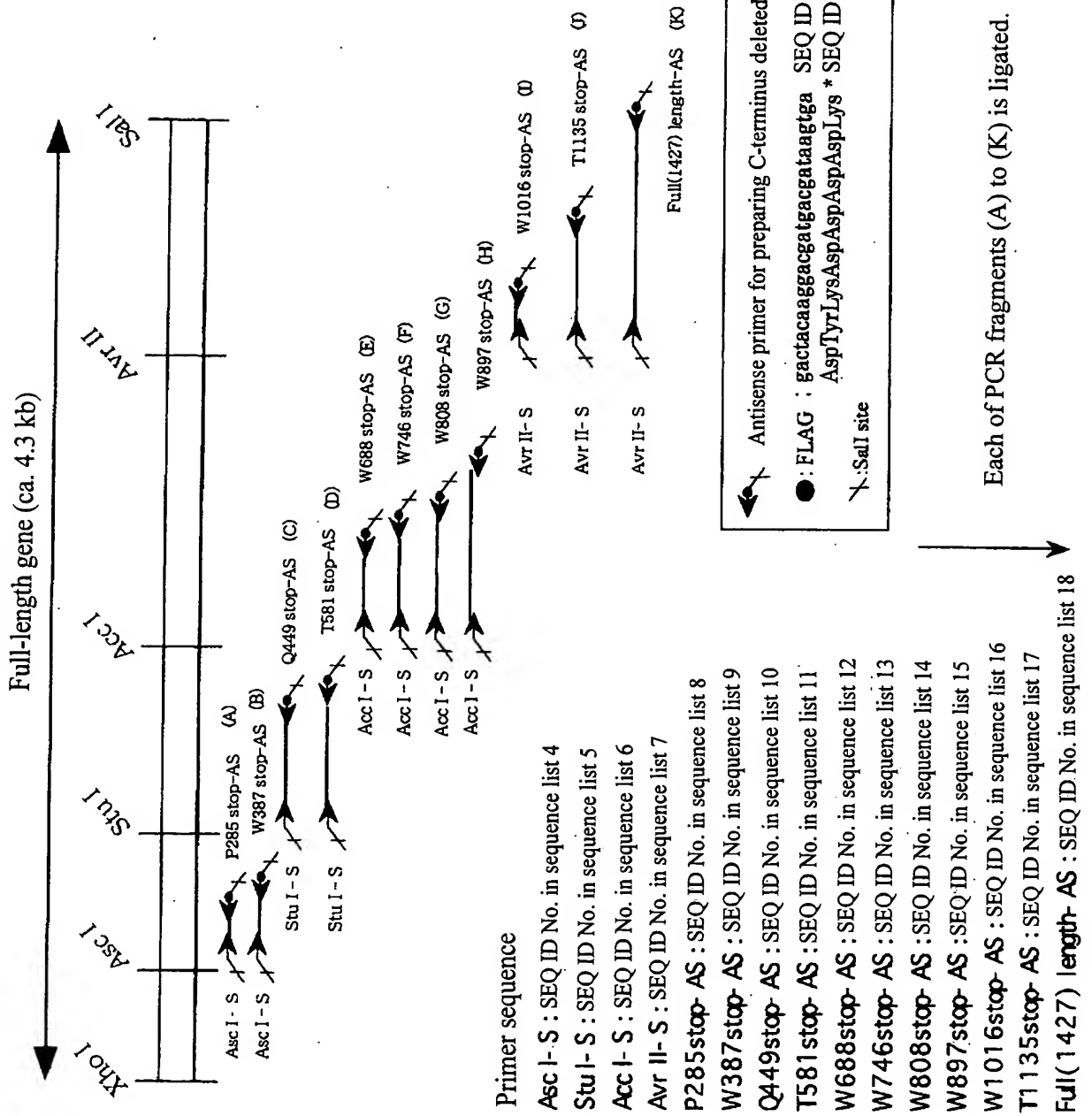
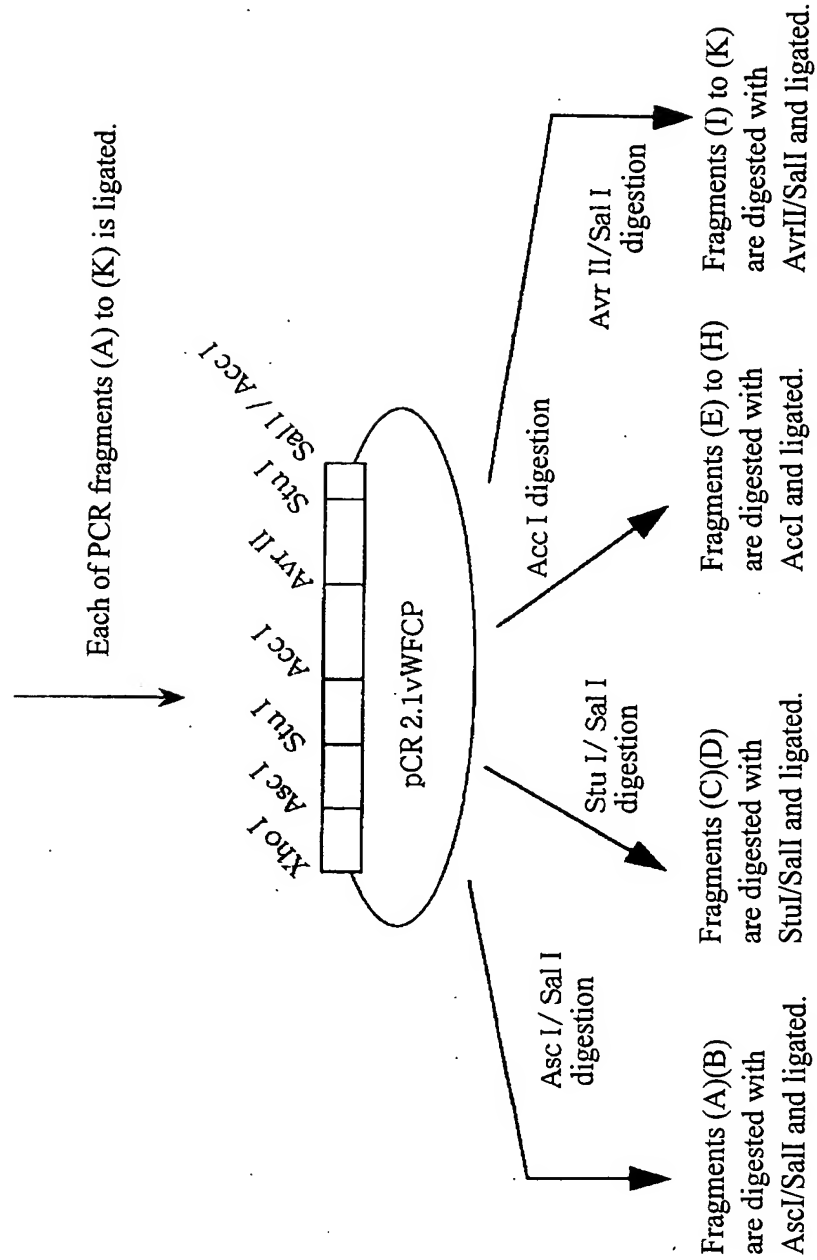


Fig. 1



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Fig. 1



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Fig. 2

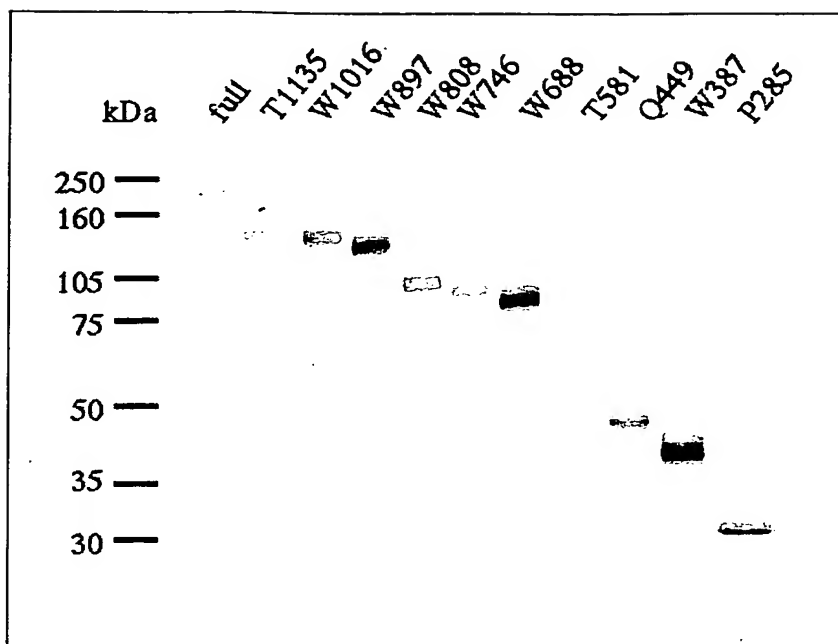
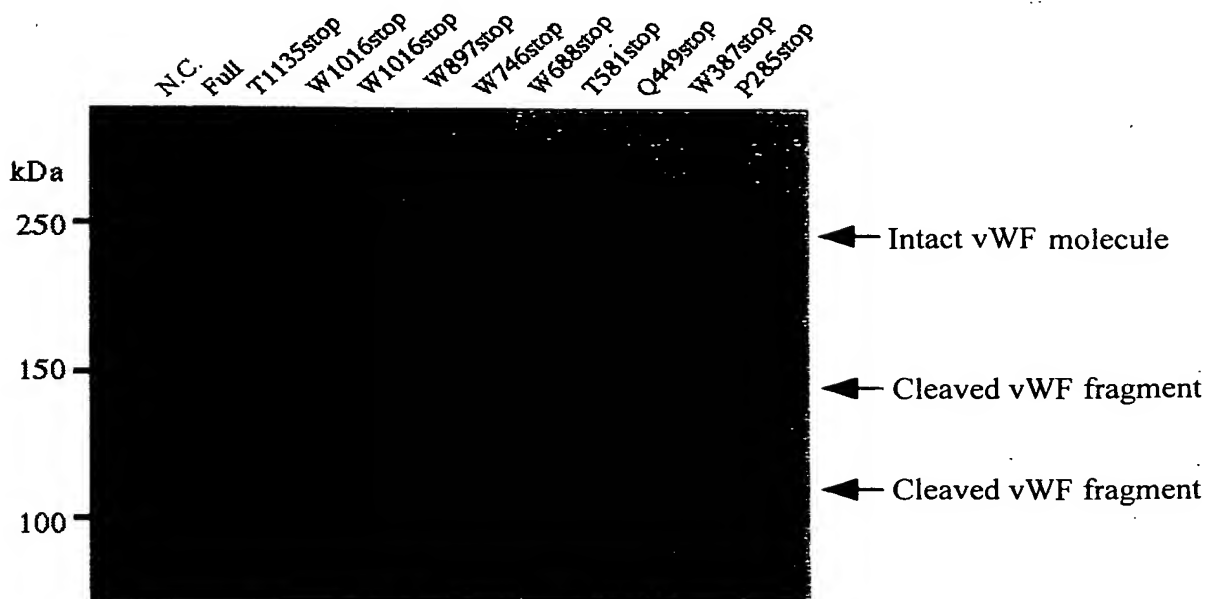


Fig. 3



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Fig. 2

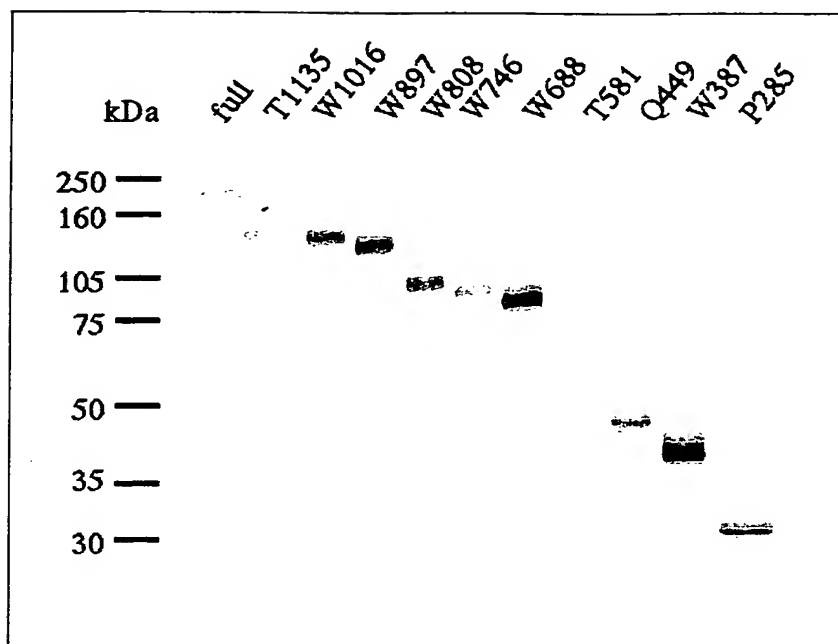
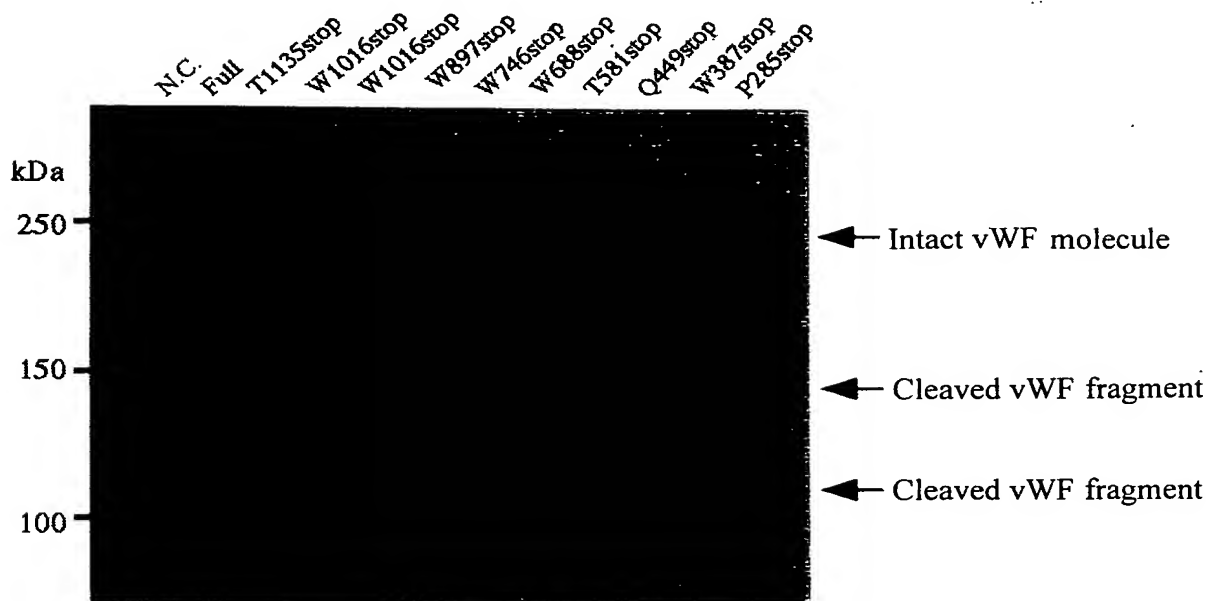


Fig. 3



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Fig. 2

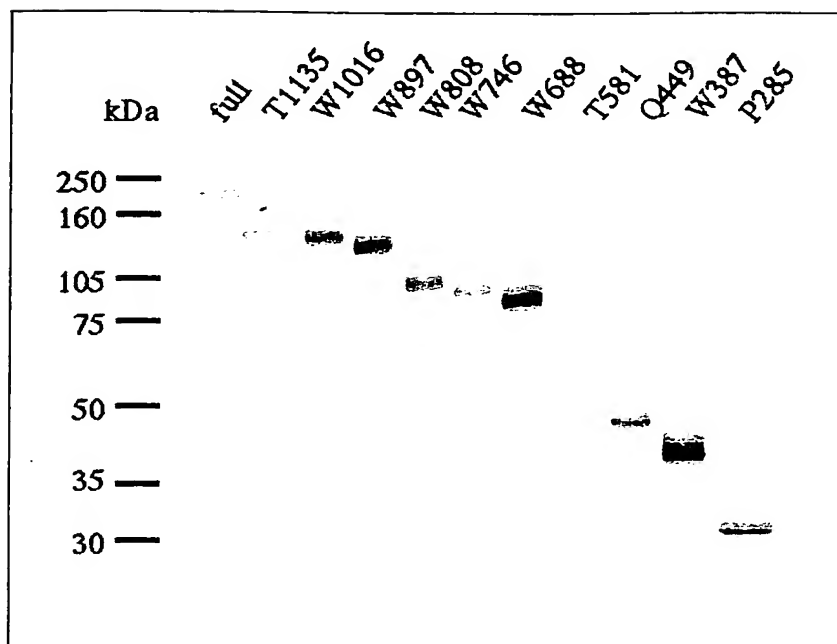
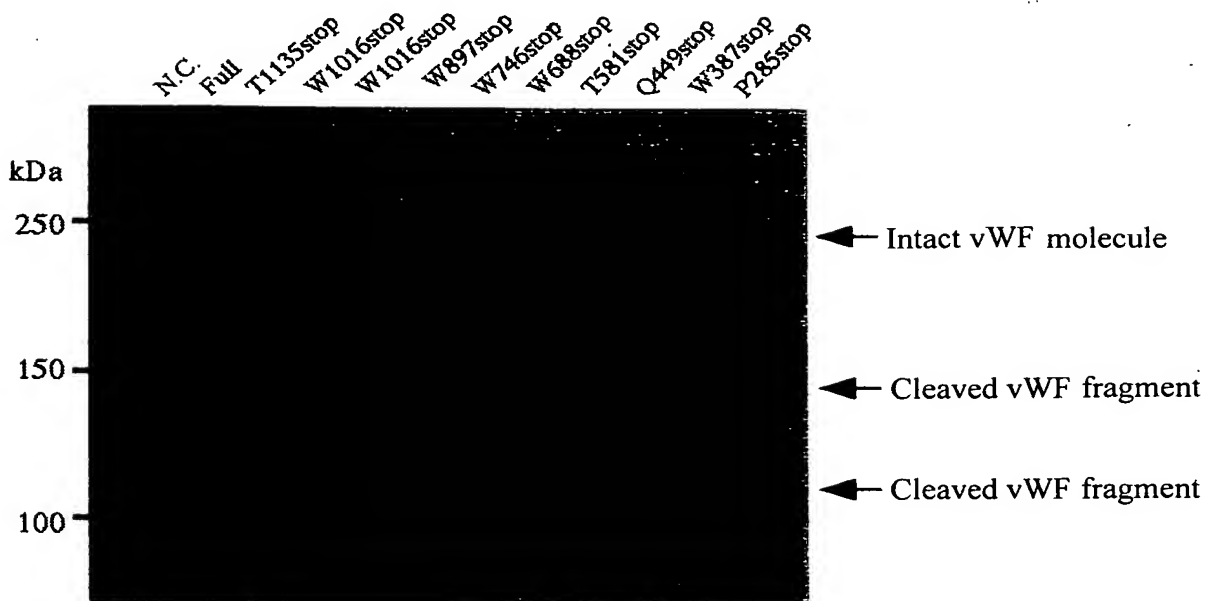


Fig. 3



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Fig. 4

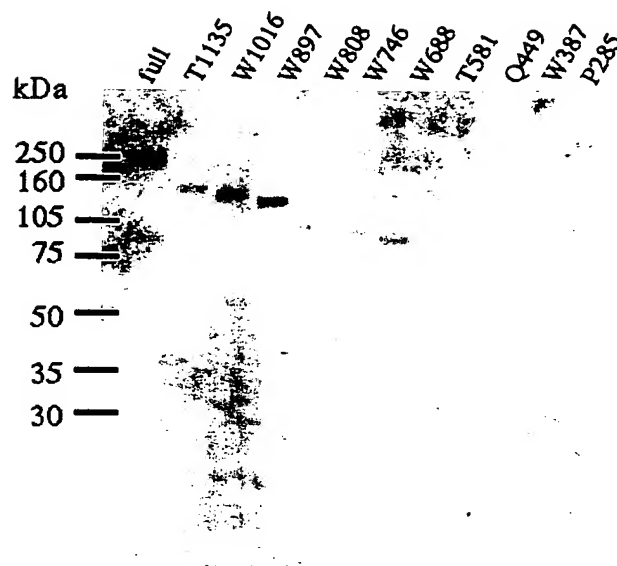
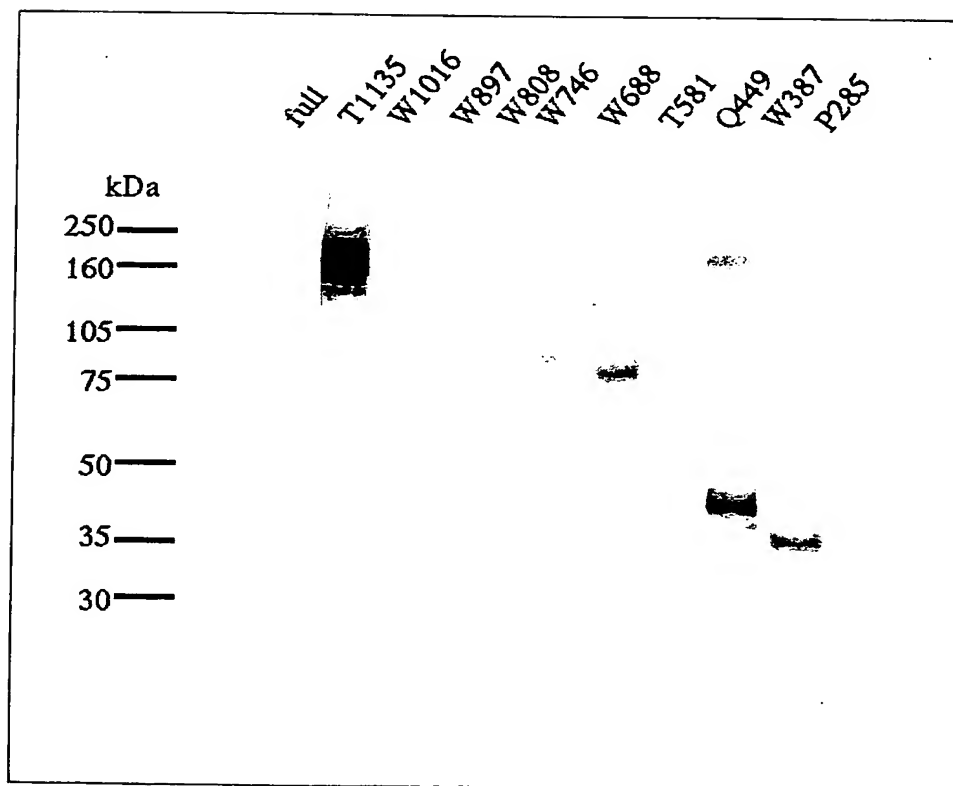


Fig. 5



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Fig. 4

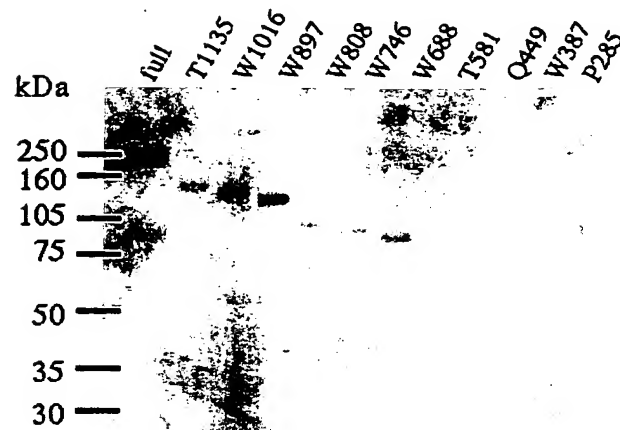
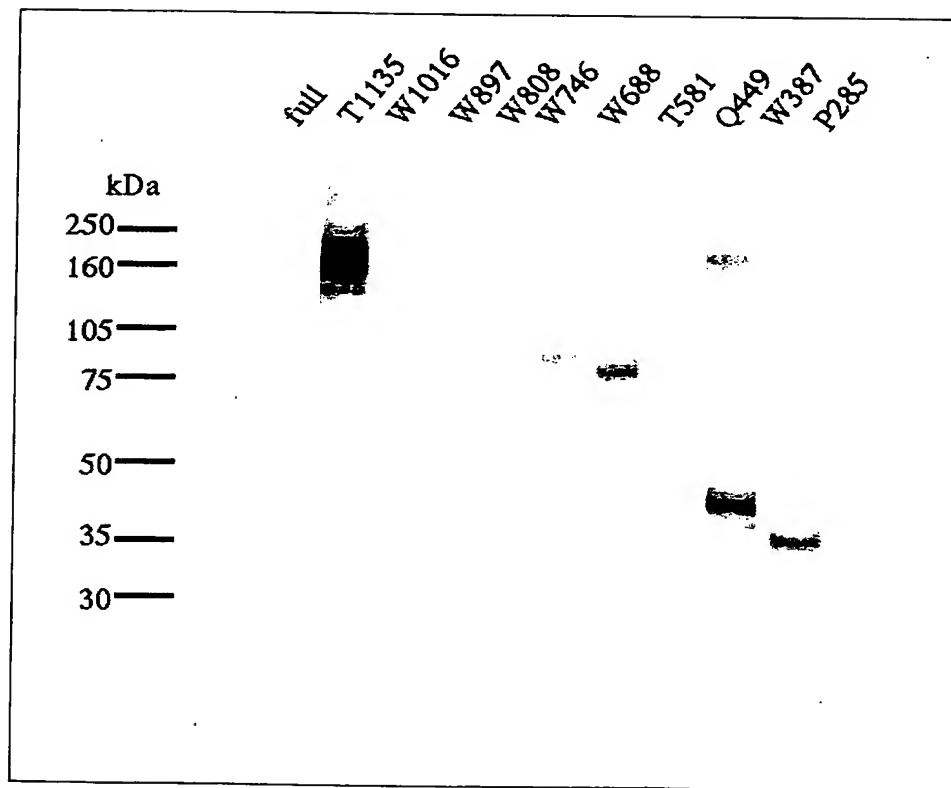


Fig. 5



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Fig. 4

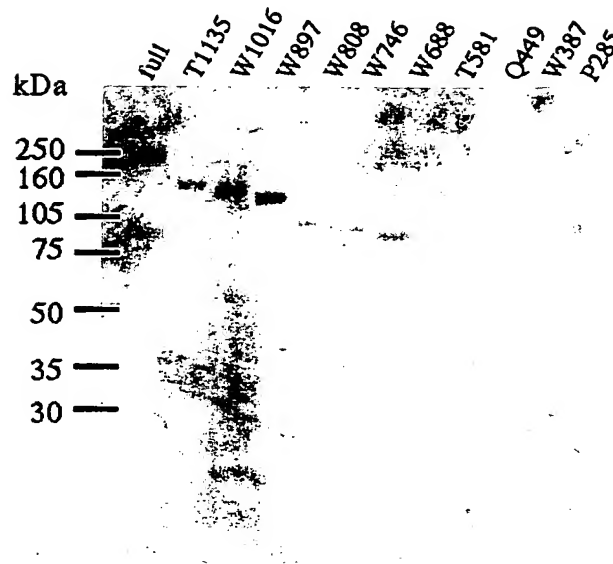
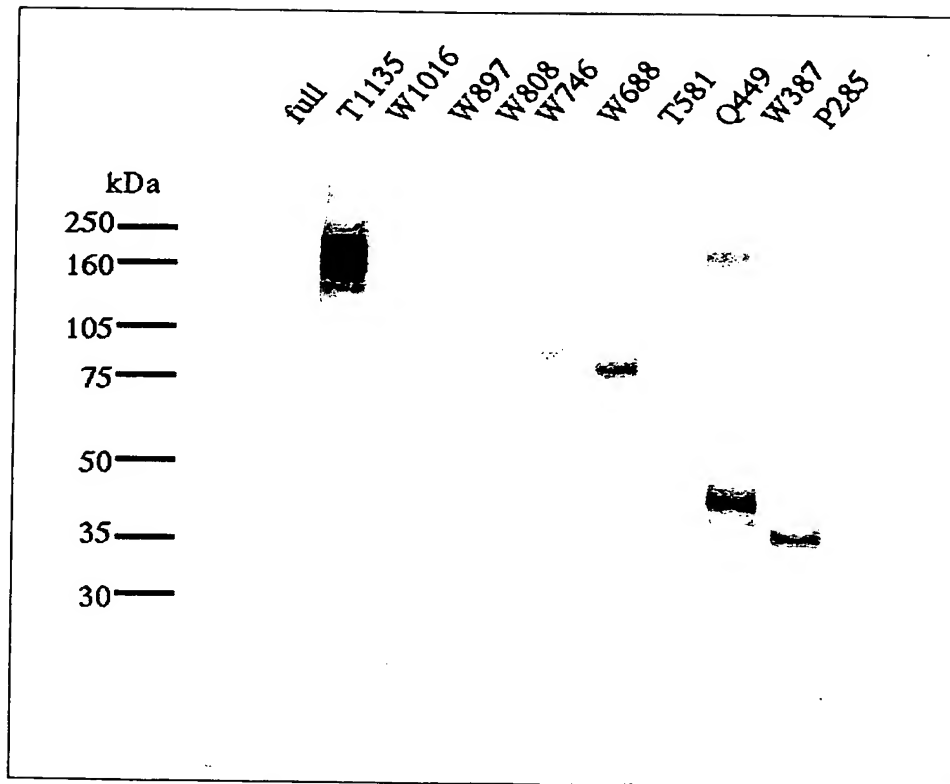


Fig. 5





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Fig. 6

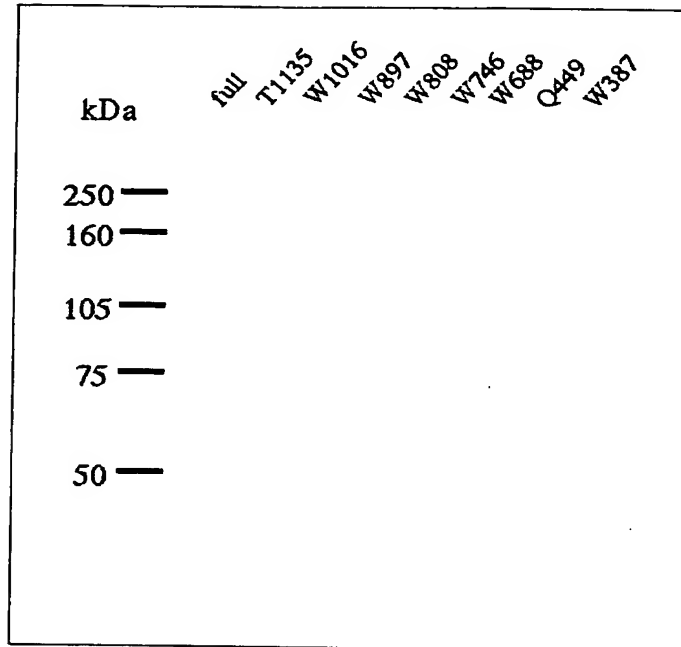
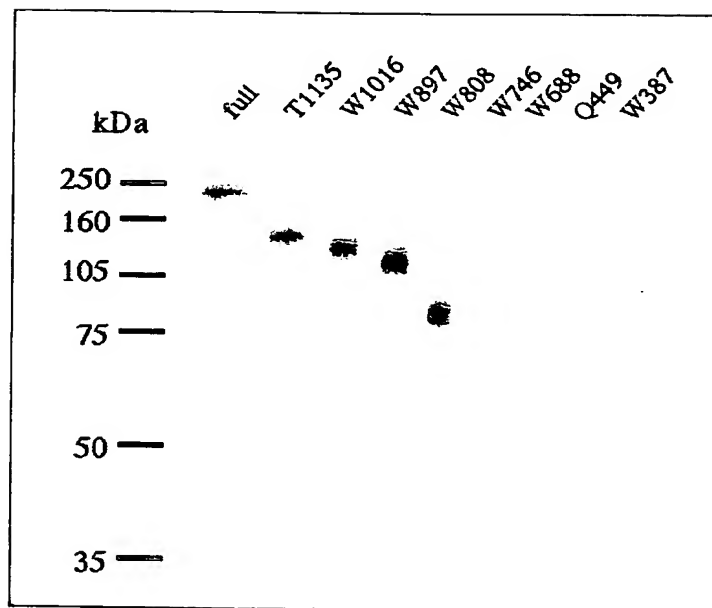


Fig. 7



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Fig. 6

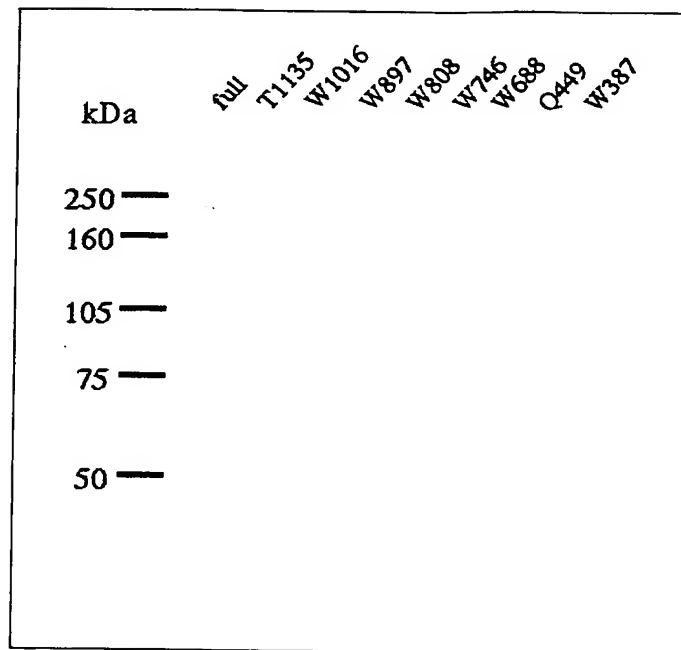
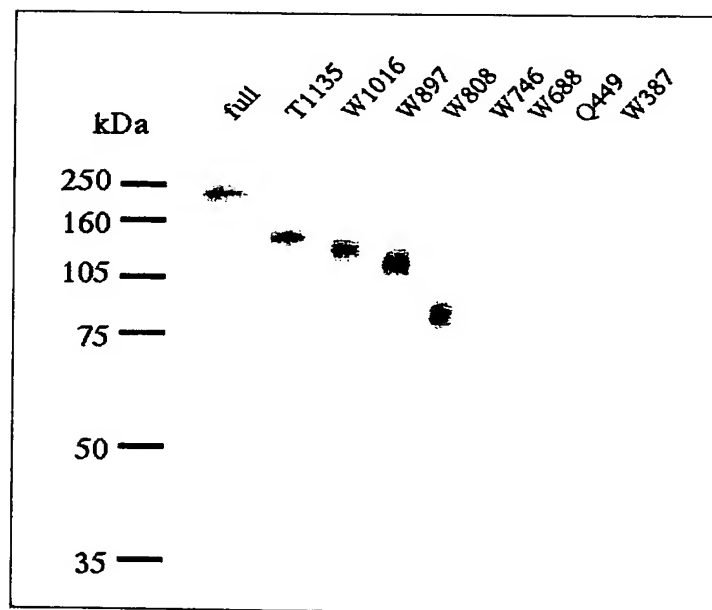


Fig. 7



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Fig. 6

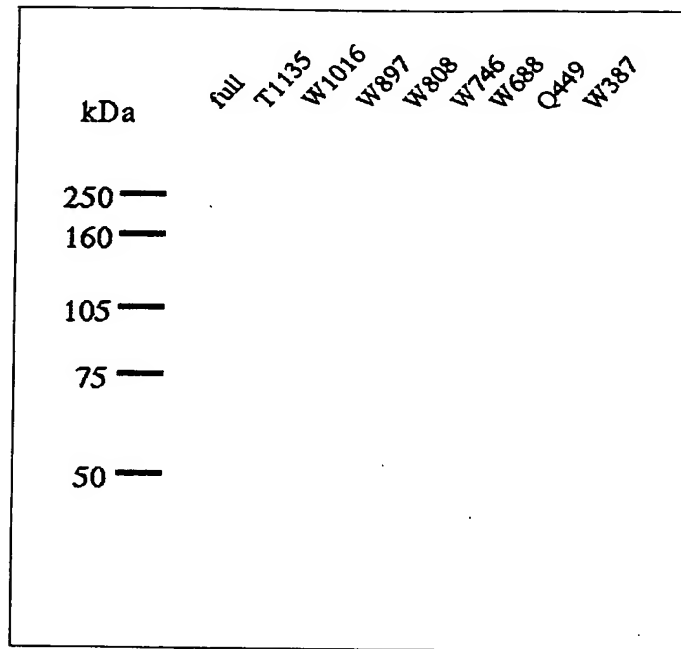
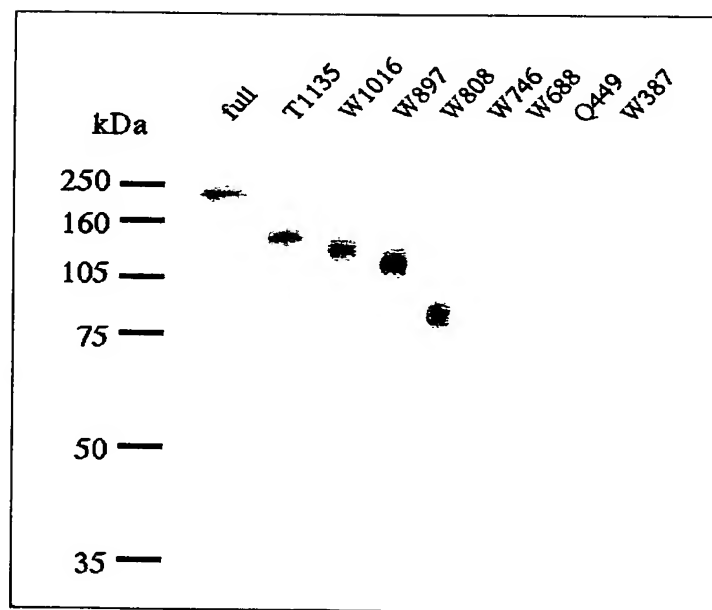


Fig. 7



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Fig. 8

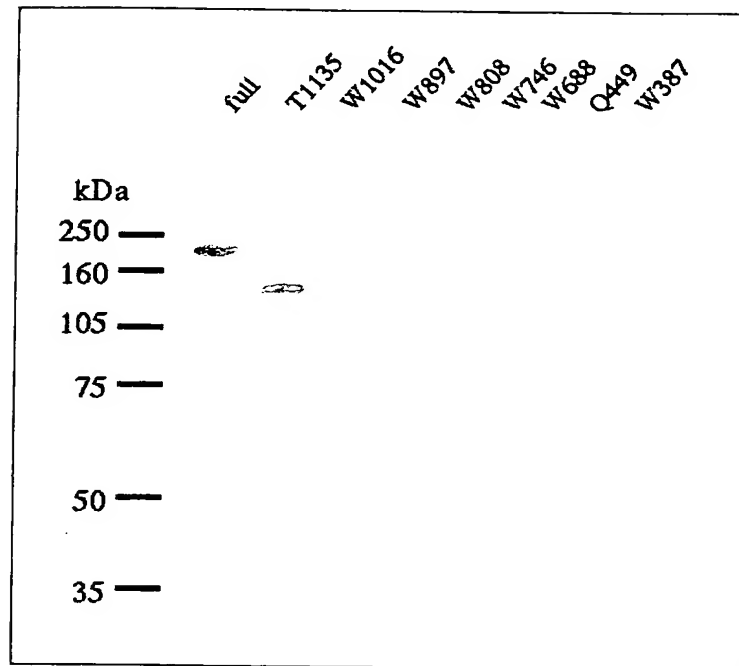
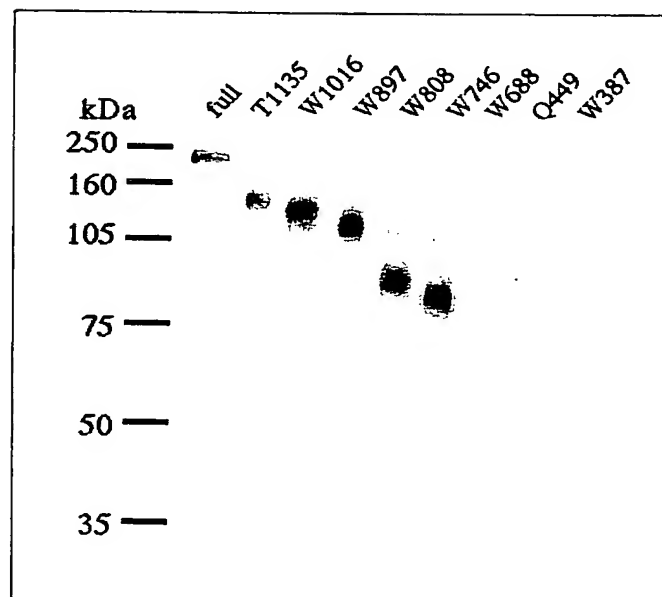


Fig. 9



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Fig. 8

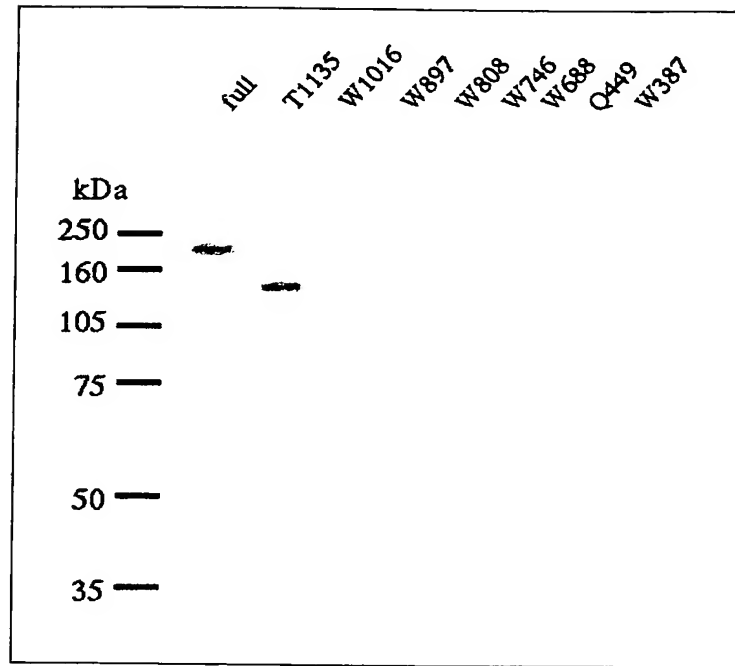
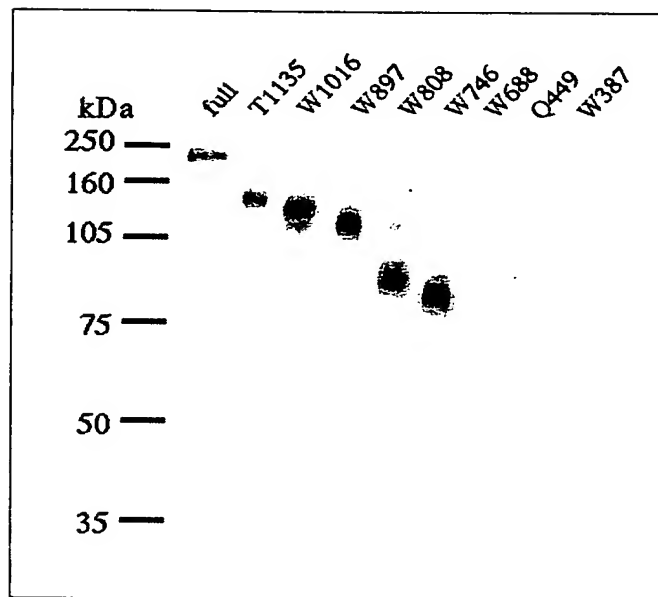


Fig. 9



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Fig. 8

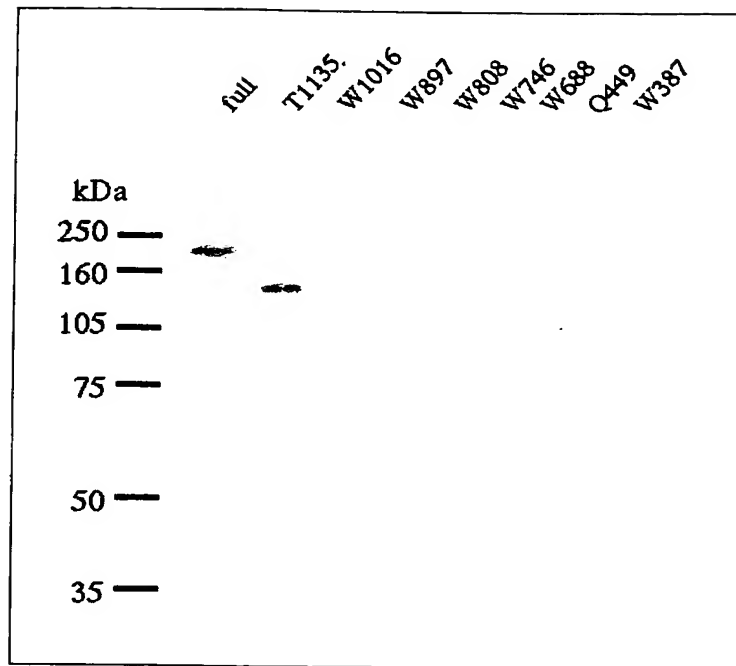
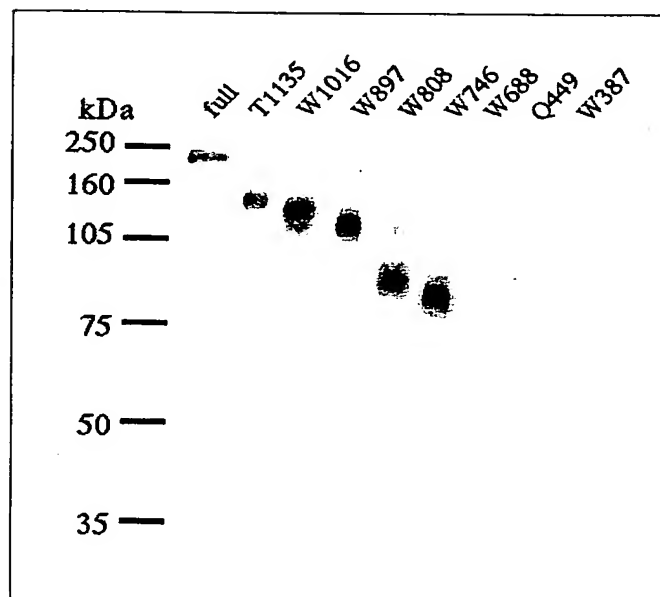


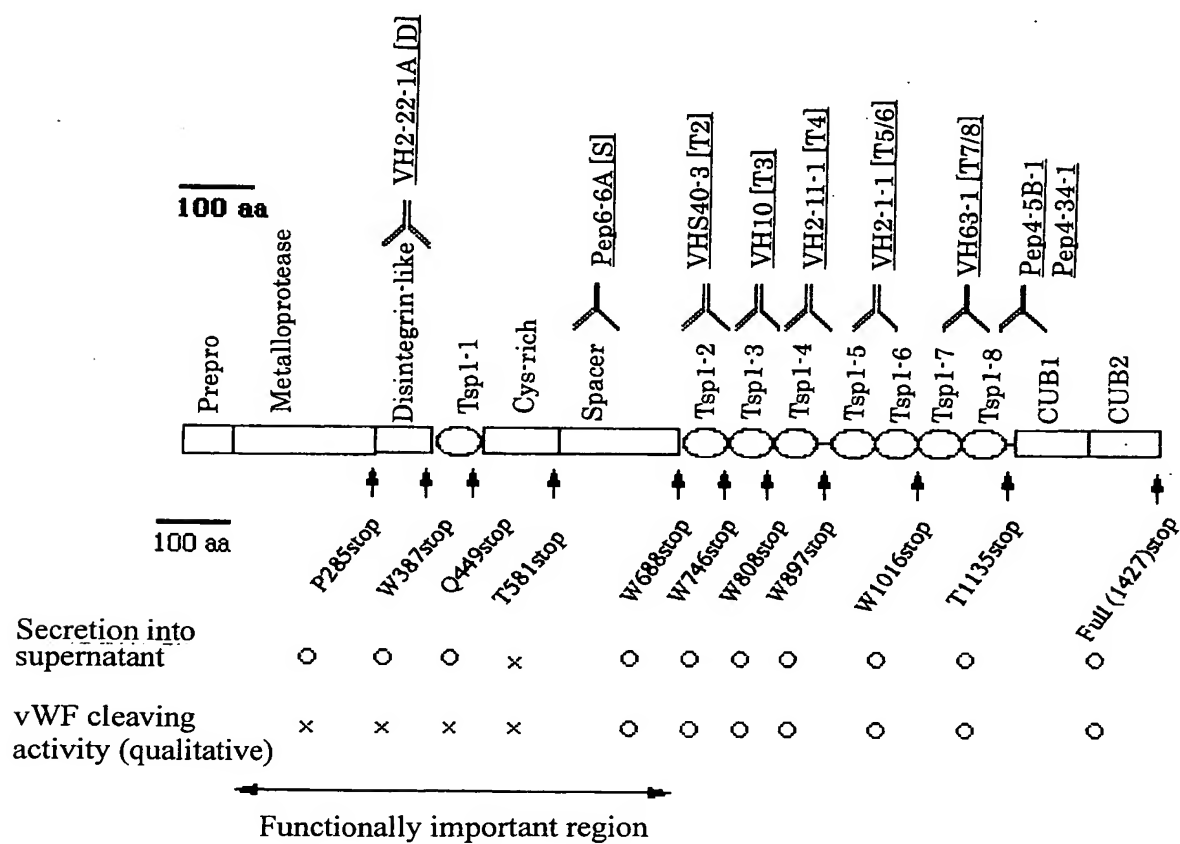
Fig. 9



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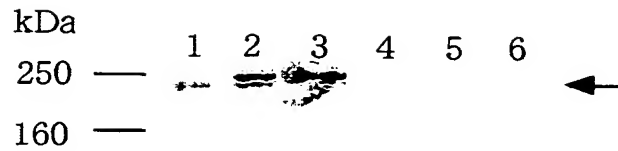
Fig. 10

- PoAb 1 Immunization region (Full-length DNA) neutralizing ability +  
 PoAb 2 Immunization region (Q449stop+FLAG DNA) neutralizing ability +  
 PoAb 3 Immunization region (P285stop+FLAG DNA) neutralizing ability +



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Fig. 11

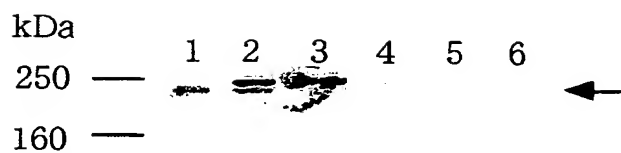


1. Gel filtration sample of FI paste of human pooled plasma
2. Plasma 1 from healthy subject
3. Plasma 2 from healthy subject
4. Plasma 3 from healthy subject
5. Plasma 1 from TTP patient
6. Plasma 2 from TTP patient



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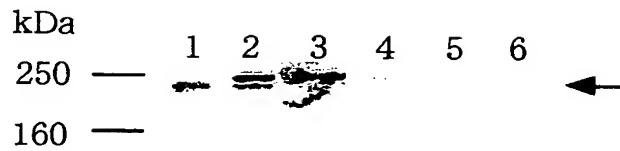
Fig. 11



1. Gel filtration sample of FI paste of human pooled plasma
2. Plasma 1 from healthy subject
3. Plasma 2 from healthy subject
4. Plasma 3 from healthy subject
5. Plasma 1 from TTP patient
6. Plasma 2 from TTP patient

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Fig. 11



1. Gel filtration sample of FI paste of human pooled plasma
2. Plasma 1 from healthy subject
3. Plasma 2 from healthy subject
4. Plasma 3 from healthy subject
5. Plasma 1 from TTP patient
6. Plasma 2 from TTP patient

Fig. 12

(MoAb-PoAb system)

1. Allow each reagent to be warmed to room temperature
2. Add a sample to WH10 MoAb immobilized plate by 100  $\mu$ L/well  
37°C, 1 hour
3. Wash the plate with 0.05% Tween-20-TBS three times
4. Dilute PoAb 1 or PoAb 2 with a diluting solution (1% BSA-TBS) so that it may become 1  $\mu$ g/ml and add to the plate by 100  $\mu$ L/well  
37°C, 1 hour
5. Wash the plate with 0.05% Tween-20-TBS three times
6. Dilute an anti-rabbit IgG-HRP labelled conjugate with a diluting solution (1% BSA-TBS) to 10000-fold and add to the plate by 100  $\mu$ L/well  
37°C, 1 hour
7. Wash the plate with 0.05% Tween-20-TBS three times
8. Add a TMB substrate solution (prepared by mixing two liquids at room temperature immediately before use) to the plate by 100  $\mu$ L/well (positive well turns blue); about 10 minutes (so that the color of 100 ng/ml of the recombinant product enclosed as a standard may be finally about 1 as OD450 nm after the reaction is terminated) at room temperature (positive well turns yellow)
9. Add a reaction terminating liquid (0.5 M sulfuric acid) to the plate by 100  $\mu$ L/well
10. Measure the plate with a plate reader at 450 nm and 650 nm

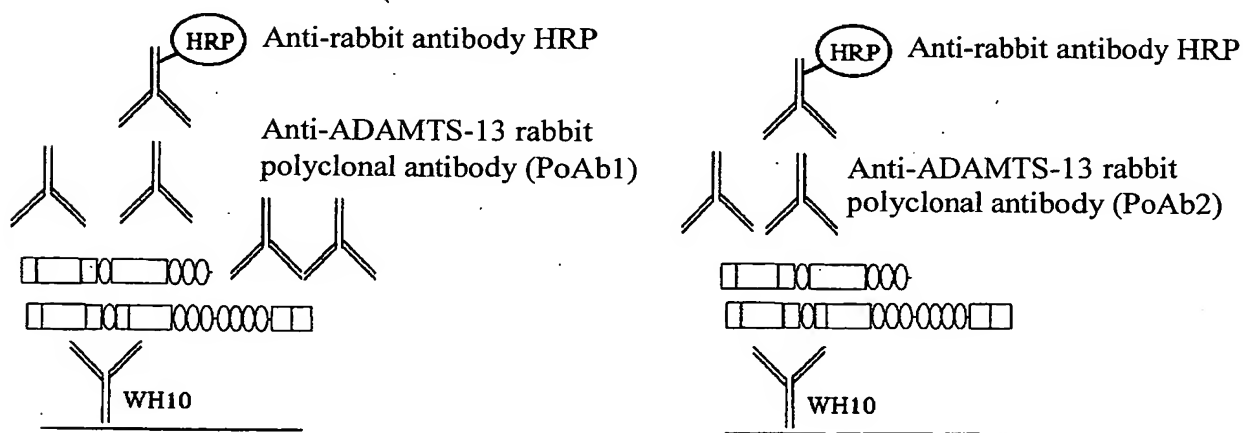
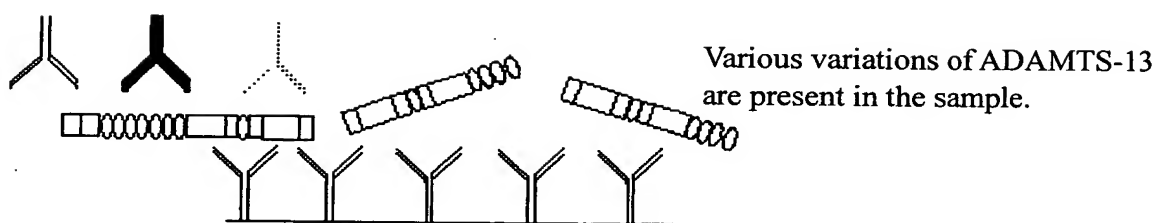


Fig. 13

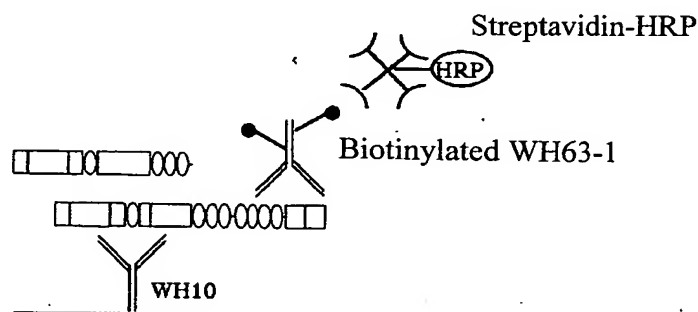


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Fig. 14

(MoAb-MoAb system)

1. Allow each reagent to be warmed to room temperature
2. Add a sample to WH10 MoAb immobilized plate by 100  $\mu$ L/well  
37°C, 1 hour
3. Wash the plate with 0.05% Tween-20-TBS three times
4. Dilute a biotinylated antibody with a diluting solution (1% BSA-TBS) so that it may become  
1  $\mu$ g/ml and add to the plate by 100  $\mu$ L/well  
37°C, 1 hour
5. Wash the plate with 0.05% Tween-20-TBS three times
6. Dilute a streptavidin-HRP labelled conjugate with a diluting solution (1% BSA-TBS)  
to 10000-fold and add to the plate by 100  $\mu$ L/well  
37°C, 1 hour
7. Wash the plate with 0.05% Tween-20-TBS three times
8. Add a TMB substrate solution (prepared by mixing two liquids at room temperature  
immediately before use) to the plate by 100  $\mu$ L/well (positive well turns blue); about 10  
minutes (so that the color of 100 ng/ml of the recombinant product enclosed as a standard  
may be finally about 1 as OD450 nm after the reaction is terminated) at room temperature  
(positive well turns yellow)
9. Add a reaction terminating liquid (0.5 M sulfuric acid) to the plate by 100  $\mu$ L/well
10. Measure the plate with a plate reader at 450 nm and 650 nm.



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Fig. 15

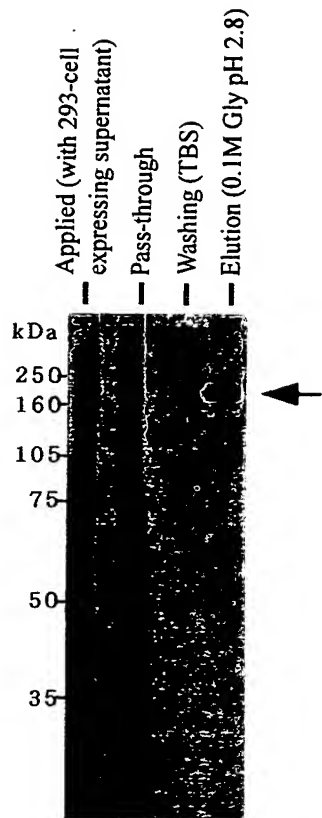
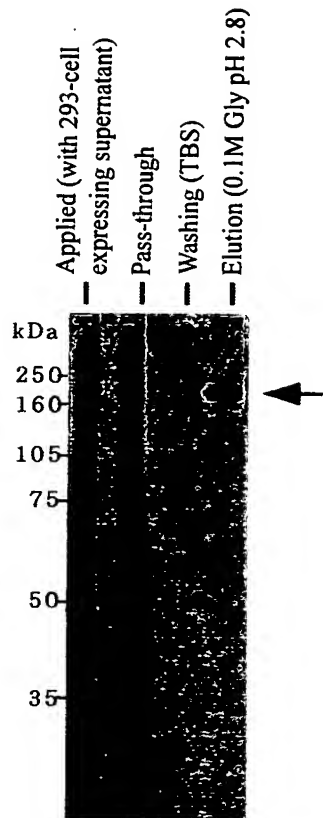
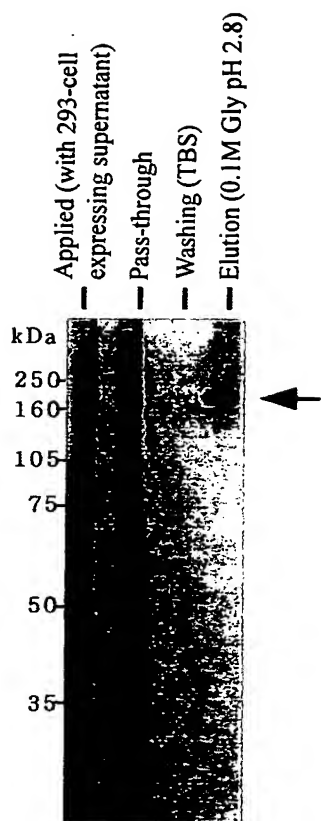


Fig. 15



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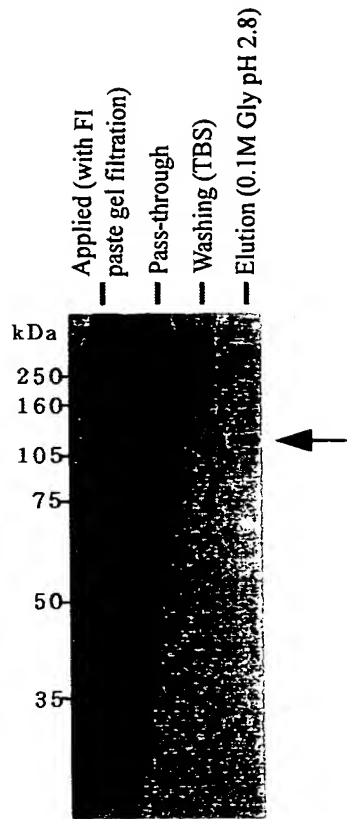
Fig. 15





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Fig. 16



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Fig. 16

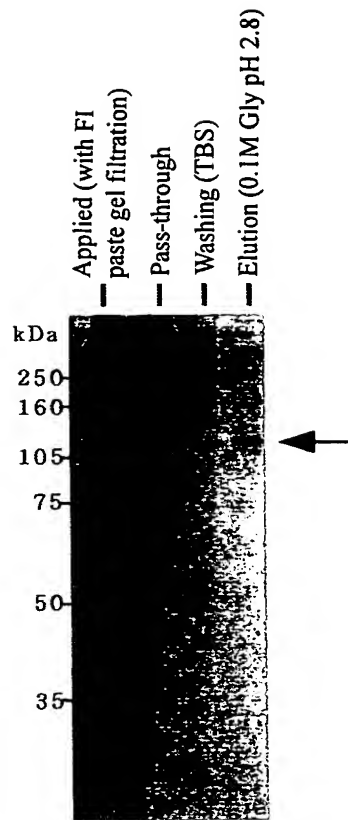
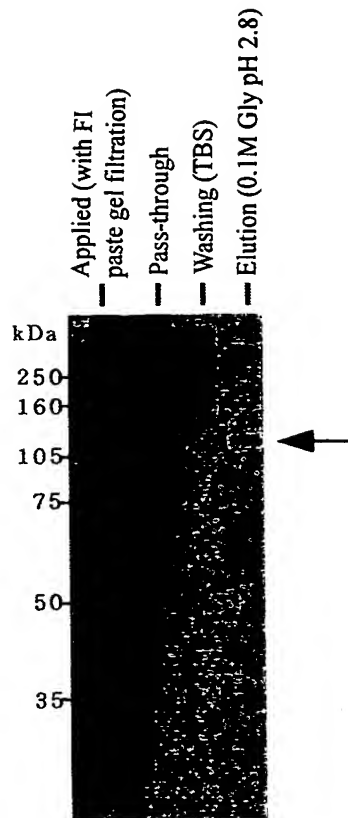
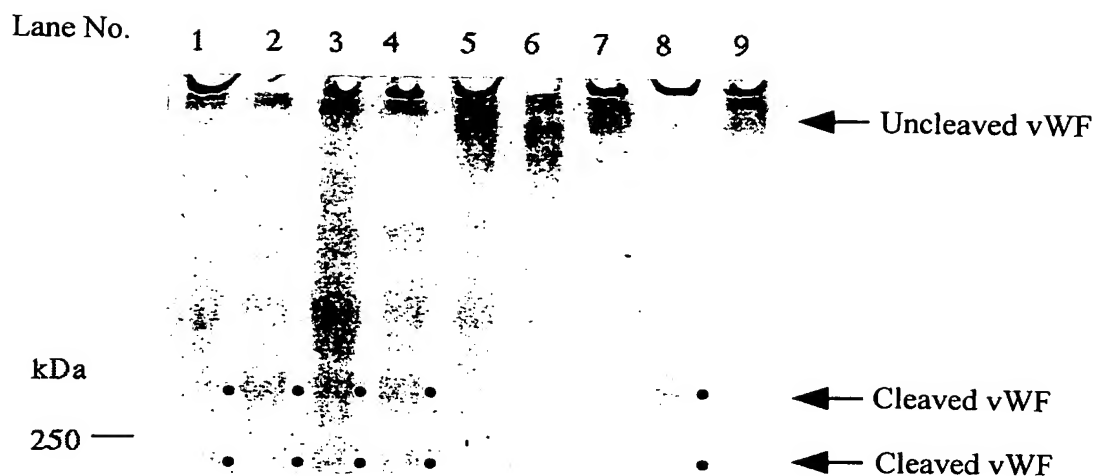


Fig. 16



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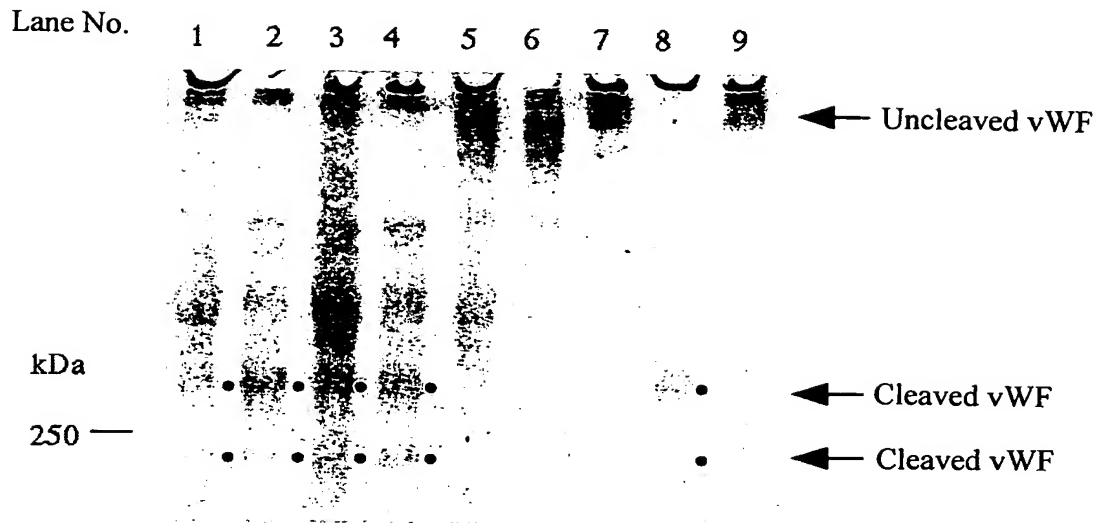
Fig. 17



1. ADAMTS-13 solution: normal rabbit serum = 1:1
2. ADAMTS-13 solution: normal rabbit serum (diluted 5-fold)= 1:1
3. ADAMTS-13 solution: peptide immunized rabbit serum = 1:1
4. ADAMTS-13 solution: peptide immunized rabbit serum (diluted 5-fold)= 1:1
5. ADAMTS-13 solution: recombinant protein immunized rabbit serum = 1:1
6. ADAMTS-13 solution: recombinant protein immunized rabbit serum (diluted 5-fold)= 1:1
7. ADAMTS-13 solution: 10 mM EDTA = 1:1
8. ADAMTS-13 solution: buffer only = 1:1
9. Buffer (without ADAMTS-13): buffer = 1:1

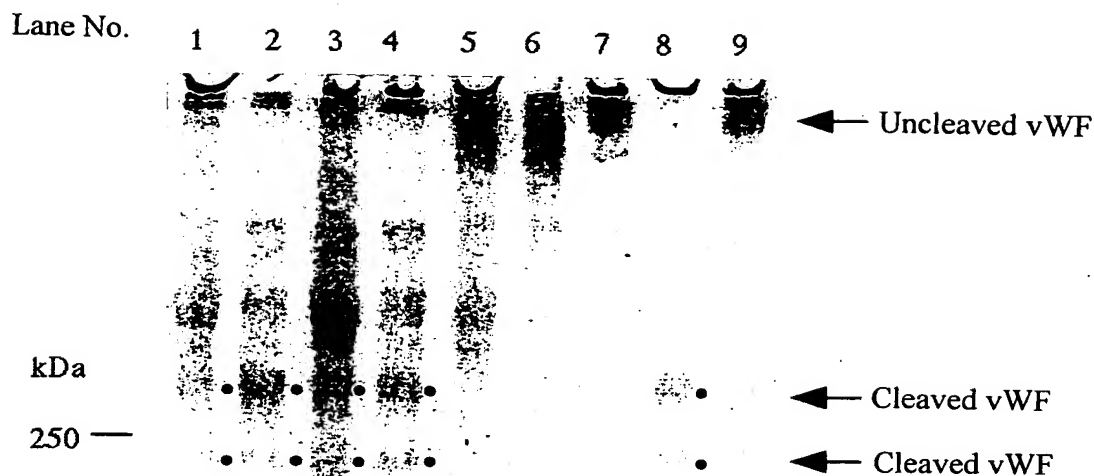
10/529009

Fig. 17



1. ADAMTS-13 solution: normal rabbit serum = 1:1
2. ADAMTS-13 solution: normal rabbit serum (diluted 5-fold)= 1:1
3. ADAMTS-13 solution: peptide immunized rabbit serum = 1:1
4. ADAMTS-13 solution: peptide immunized rabbit serum (diluted 5-fold)= 1:1
5. ADAMTS-13 solution: recombinant protein immunized rabbit serum = 1:1
6. ADAMTS-13 solution: recombinant protein immunized rabbit serum (diluted 5-fold)= 1:1
7. ADAMTS-13 solution: 10 mM EDTA = 1:1
8. ADAMTS-13 solution: buffer only = 1:1
9. Buffer (without ADAMTS-13): buffer = 1:1

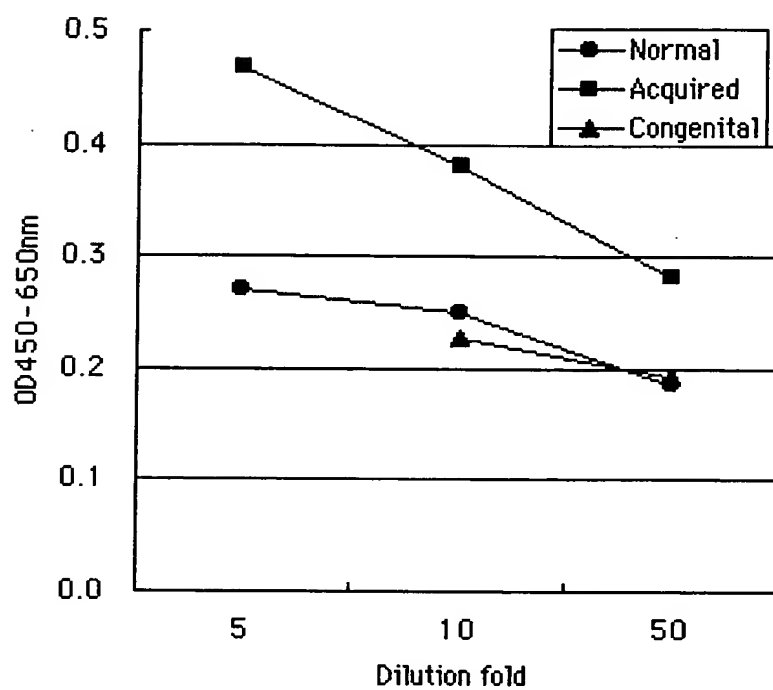
Fig. 17



1. ADAMTS-13 solution: normal rabbit serum = 1:1
2. ADAMTS-13 solution: normal rabbit serum (diluted 5-fold)= 1:1
3. ADAMTS-13 solution: peptide immunized rabbit serum = 1:1
4. ADAMTS-13 solution: peptide immunized rabbit serum (diluted 5-fold)= 1:1
5. ADAMTS-13 solution: recombinant protein immunized rabbit serum = 1:1
6. ADAMTS-13 solution: recombinant protein immunized rabbit serum (diluted 5-fold)= 1:1
7. ADAMTS-13 solution: 10 mM EDTA = 1:1
8. ADAMTS-13 solution: buffer only = 1:1
9. Buffer (without ADAMTS-13): buffer = 1:1

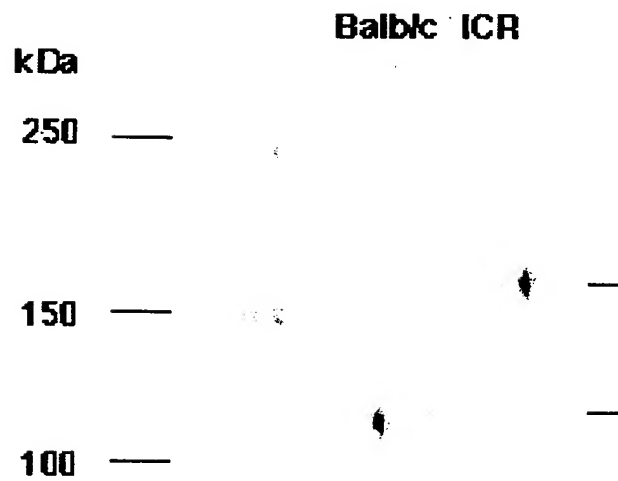
10/529009

Fig. 18



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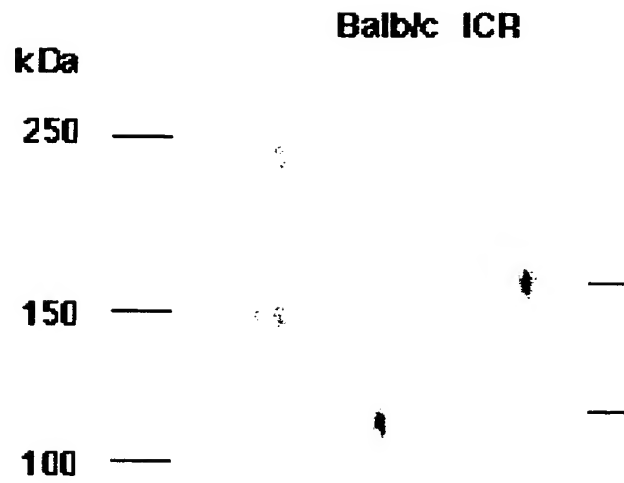
Fig. 19





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Fig. 19



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FIG. 19: 10/529009

Fig. 19

